CREATING THE NEXT®

Georgia

Ribonuclease H2A (RNASEH2A) is the catalytic subunit of RNASEH2, a key enzyme for removal of incorporated ribonucleotides in DNA and maintenance of genomic integrity for normal cellular growth and division. Gene Ontology analysis on genes showing high positive correlation with RNASEH2A in mitotic cell cycle regulation. Analysis of expression correlations was performed on a list of genes containing the RNASEH2A, RNASEH2A patient samples from 35 cancer types. Clustering of expression correlation. The analysis revealed positive correlation of RNASEH2A with genes highly expressed in cancer and cancer proliferation markers as compared to RNASEH2A expression is highly associated with specific cell cycle markers, i.e., G2 and M cell cycle markers as opposed to G1 and S cell cycle markers. The observed correlations were successfully validated using the Broad institute's Cancer Cell Line Encyclopedia (CCLE) containing ~1000 cell lines. Copy Number Alteration in TCGA Pan Cancer dataset showed high prevalence of RNASEH2A gene amplification in multiple cancer types, further providing genomic evidence of elevated RNASEH2A expression in cancer. Our bioinformatic study, clearly shows increased co-expression of RNASEH2A with cancer proliferation and mitotic cell cycle markers, suggesting that RNASEH2A expression level could be a predictor of poor outcomes and a possible target for therapeutic interventions.

INTRODUCTION

Differential cell-cycle regulation of the RNASEH2A orthologous gene has been observed in yeast S. Cerevisiae and increase in levels of RNASEH2A has previously been observed with overexpression of several oncogenes in mesenchymal stem cells.

Study of co-expressed genes with RNASEH2A can reveal other genes involved with RNASEH2A either in common or interconnected biological processes.

Copy Number Alterations(CNAs) are somatic changes to chromosome structure that result in gain or loss in copies of sections of DNA

Classification of CNAs

- Deep deletion: deep loss/homozygous deletion;
- Shallow deletion: shallow loss/heterozygous deletion
- Diploid: homozygous genes
- Gain: a low-level gain/additional copies
- Amplification: high-level amplification

Deep deletions and **Amplification** causes highest burden, prognostic for cancer specific death.



- . Find suggestive role of RNASEH2A in human tissues by using RNASEH2A co-expressed genes for gene ontology analysis
- 2. To elucidate the role of RNASEH2A in different cell cycle phases and in cancer via expression correlation analysis with marker genes
- 3. To validate RNASEH2A elevated expression in cancer by measuring the prevalence of CNAs of RNASEH2A in different cancer subtypes

MATERIAL (DATASETS)

GTEx

TCGA∉∋

CBioPortal FOR CANCER GENOMICS

1. RNA-seq expression data in Transcripts per million (TPM) from **Genotype-Tissue Expression (GTEx)** portal v7 is available for 53 different human tissues



3. Copy number alterations (CNAs) and RNA-Seq data from **<u>The</u>** Cancer Genome Atlas (TCGA) Pan **Cancer studies** involving 32 studies and 10,967 patients

different tissues of origin



were used for Gene Ontology analysis

Process				
GO Term	Description	<i>p</i> -Value	FDR <i>q</i> -Value	
GO 1903047	mitotic cell cycle process	$2.80 imes 10^{-14}$	$5.93 imes 10^{-11}$	
GO 0070507	regulation of microtubule cytoskeleton organ	$1.39 imes 10^{-7}$	$8.40 imes10^{-5}$	
GO 0006974	cellular response to DNA damage stimulus	2.77×10^{-5}	$5.86 imes 10^{-3}$	
GO 0007059	chromosome segregation	$9.00 imes 10^{-5}$	$1.36 imes 10^{-2}$	
GO 0006260	DNA replication	$2.66 imes 10^{-4}$	3.12×10^{-2}	
GC 0000200	DINA replication	2.00×10	3.12×10^{-1}	

1.b. Gene Ontology(GO) term analysis of top 2% co-expressed genes with RNASEH2A in GTEx dataset reveals **possible involvement in Mitotic <u>Cell Cycle Process</u>**

Cancer Subtype(s)	Prevalence	Prevalence of Copy Number Alterations(CNAs) of RNASEH2A gene				Average RNASEH2A mRNA expression each CNA group, RSEM					
	Deep Deletion	Shallow Deletion	Diploid	Gain	Amplification		Deep Deletion	Shallow Deletion	Diploid	Gain	Amplificati
Ovarian Epithelial Tumor	0.34%	29.15%	26.10%	36.27%	8.14%		649.61	730.15	1150.31	1444.92	2435.81
Endometrial Carcinoma		12.13%	69.67%	14.90%	3.29%			838.32	912.12	1721.50	2087.09
Adrenocortical Carcinoma		1.32%	34.21%	61.84%	2.63%			707.14	480.63	795.96	1114.81
Pleural Mesothelioma		8.05%	73.56%	16.09%	2.30%			631.88	534.82	975.79	1203.17
Esophageal Squamous Cell Carcinoma		34.04%	44.68%	19.15%	2.13%			775.80	697.66	974.64	2097.29
Cervical Squamous Cell Carcinoma	0.41%	26.23%	57.79%	13.52%	2.05%		1700.49	1289.19	1556.32	2079.49	7219.93
Diffuse Glioma	0.20%	3.33%	73.92%	20.78%	1.76%		205.72	413.52	351.50	461.54	404.56
Sarcoma		10.36%	50.20%	37.85%	1.59%			812.55	808.58	1356.59	1326.82
Invasive Breast Carcinoma	0.09%	21.25%	60.11%	17.23%	1.31%		384.40	618.31	615.99	940.40	1485.75
Ocular Melanoma		3.75%	92.50%	2.50%	1.25%			731.66	617.38	1234.06	798.60
3. Prevalence and average mRNA expression in patient samples with different Copy number variations for RNASEH2A gene in cancer subtypes. Average expression increas patients found with amplified copy of RNASEH2A gene in comparison to average expression of patients with diploid RNASEH2A gene subtype. Prevalence of Amplificati											

Expression correlation and Copy Number Alteration analysis of RNASEH2A in cancer datasets confirms its association with cancer proliferation and specific cell cycle markers Deepali L. Kundnani, Stefania Marsili, Ailone Tichon and Francesca Storici School of Biological Sciences, Georgia Institute of Technology, Atlanta, GA, USA

ABSTRACT

METHODOLOGY

The above list contains 1. Markers involved/associated in different cell cycle phases, Markers up/down regulated in cancer, 3. Cell cycle associated cancer markers, 4. Genes of our interest(RNASEH1, subunits of RNASEH2)

1.a. RNASEH2A gene expression is seen to be **highest in Testis (~50 TPM)**, which is more than 2X of RNASEH2A expression in any other human tissue in GTEx dataset



2. Correlation Plots of selected genes in TCGA Pan Cancer(i) and CCLE(ii) Datasets with hierarchical clustering to reveal association of RNASEH2A (in red) (as opposed to RNASEH2B, RNASEH2C and RNASEH1) with cancer Proliferation and G2 <u>Cell cycle markers</u>

<u>RNASEH2A gene copy is seen to be higher than prevalence of Deep Deletion in many cancer subtypes.</u>

/Role	References	
e Phase	A. Subramanian et al. (GSEA Database)	
ele Phase	A. Subramanian et al. (GSEA Database)	
e Phase	A. Subramanian et al. (GSEA Database)	
e Phase	A. Subramanian et al. (GSEA Database)	
cle Phase	A. Subramanian et al. (GSEA Database)	
e Phase	A. Subramanian et al. (GSEA Database)	
cle Phase	A. Subramanian et al. (GSEA Database)	
n cancer	M. Li et al	
in cancer	M. Li et al	
ers in cancer	M. L. Whitfield et al.	
associated with	M. L. Whitfield et al.	
62 and M)		
f interest	This study	

2.b Expression correlation analys **TCGA and CCLE expression dataset**

Pearson Correlation analysis was between each pair 40 selected followed by clustering to reorder with similar expression trends and degree of association

3. Prevalence of Copy Nu Alterations(CNAs) in TCGA Pan Ca data

Prevalence for each CNA type calculated in patients from each ca 2. subtype

	CONCLUSION
sis in ts done jenes,	1. GO analysis of RNASEH2A highly co-expressed genes in human tissues and correlation analysis in CCLE and TCGA concludes <u>RNASEH2A</u> expression association with mitotic cell cycle
genes high	2 High positive correlation of RNASEH2A with
mber ancer	markers unregulated in cancer and cancer proliferation markers suggests RNASEH2A plays <u>a role in cancer proliferation</u>
was ancer	3. Higher prevalence of RNASEH2A amplification vs Deep Deletion in various cancer subtypes support and validate <u>high level of RNASEH2A</u> <u>and being associated with poor prognosis in</u> <u>multiple cancer types.</u>
	<u>REFERENCES</u>
n's R	1. Marsili, S.; Tichon, A.; Kundnani, D.; Storici, F. Gene Co- Expression Analysis of Human RNASEH2A Reveals Functional Networks Associated with DNA Replication, DNA Damage Response, and Cell Cycle
1.0	Regulation. Biology 2021, 10, 221. 2 Reijns MA: Rabe B: Rigby RF: Mill P: Astell KR:
0.8	Lettice, L.A.; Boyle, S.; Leitch, A.; Keighren, M.; Kilanowski, F.; et al. Enzymatic removal of
0.6	genome integrity and development. Cell 2012, 149, 1008–
0.4	 3. GTEx Consortium. The genotype-tissue expression (GTEx) project. Nat. Genet. 2013, 45, 580-585.
0.2	4. Liu, J.; Lichtenberg, T.; Hoadley, K.A.; Poisson, L.M.; Lazar, A.J.; Cherniack, A.D.; Kovatich, A.J.; Benz, C.C.; Levine, D.A.; Lee, A.V.; et al. An Integrated TCGA Pan-
- 0.2	Cancer Clinical Data Resource to Drive High-Quality Survival Outcome Analytics. Cell 2018, 173, 400–416.e11. 5. Ghandi, M.; Huang, F.W.; Jané-Valbuena, J.; Kryukov,
- 0.4	E.T.; Bielski, C.M.; Li, H.; et al. Next-generation characterization of the Cancer Cell Line Encyclopedia.
- 0.6 - 0.8	 6. Shao, X.; Lv, N.; Liao, J.; Long, J.; Xue, R.; Ai, N.; Xu, D.; Fan, X. Copy number variation is highly correlated with differential gene expression: A pan-cancer study BMC
- 1.0 <u>close</u> and M	 Med. Genet. 2019, 20, 175. 7. Subramanian, A.; Tamayo, P.; Mootha, V.K.; Mukherjee, S.; Ebert, B.L.; Gillette, M.A.; Paulovich, A.; Pomeroy, S.L.; Golub, T.R.; Lander, E.S.; et al. Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. Proc. Natl. Acad. Sci. USA 2005. 102. 15545–15550.
ion	 Whitfield, M.L.; George, L.K.; Grant, G.D.; Perou, C.M. Common markers of proliferation. Nat. Rev. Cancer. 2006, 2, 99–106.
	ACKNOWLEDGEMENTS
es in	This research was funded by the National Institute of Health, NIH, NIGMS R01 GM115927 (F.S.); NIH NIEHS R01ES026243 (F.S.); the National Science Foundation, NSF, MCB-1615335 (F.S.) and the Howard Hughes Medical Institute Faculty Scholar grant 55108574 (F.S.).
on of	<u>CONTACT</u> Deepali Kundnani: <u>dkundnani2@catech</u> edu (D.K.)
	Francesca Storici: storici@gatech.edu (F.S.)